

STABILITY POPULATION ASSAY: METAL DISCS/METAL STRIPS/WIRES

LOT # _____ LABELED POP _____

CARRIER (circle one): Strip Disc Wire TSA Lot

ORGANISM (circle one): *B. atrophaeus* *G. stearothermophilus*
Other: _____

PROCEDURE:

- 1.0 Aseptically transfer 1 disc, strip or wire into a water blank containing 9.9 ml sterile, processed water with 0.1 ml of Tween 80 and 1ml of 3mm sterile glass beads. Vortex for 2 minutes. Place in refrigerator overnight.
2. Remove 10 ml tube from refrigerator and vortex for 2 minutes. Insert into sonicator (38.5 – 40.5 KHz, full wave industrial stack transducer) for 10 minutes.
- 3.0 Heat shock tubes in a water bath (10 minutes at 80° - 85°C for *B. atrophaeus*, 15 minutes at 95° 100°C for *G. stearothermophilus*.) Immediately cool tubes in a water bath of 0° - 4°C.

Start Time/Temperature: _____ / _____ °C End Time: _____

Initial and Date: _____ / _____

- 4.0 Vortex the tube for 1 minute. **IMPORTANT:** Make sure that metal carrier does not get stuck in the tube during vortexing. The carrier **must** flow freely with the glass bead. **Prior to performing serial dilutions, visually check to make sure that the spore deposit has been completely removed from the carrier.**
- 5.0 Perform serial dilutions by pipetting out 1.0 ml of the aliquot into another sterile, screwcapped 10 ml test tube containing 9.0 ml of sterile, processed water. Repeat from step 3 until desired dilution factor is reached.
- 6.0 From the next-to-the-last dilution, pipette out 1.0 ml into each of three petri plates. Repeat for final dilution.
- 7.0 Within 20 minutes, add to each plate approximately 20 ml of TSA, pre-sterilized and cooled to 47° ± 2°C. Swirl to distribute spores evenly in agar and allow to solidify.

TSA Temperature: _____ °C Initial and Date: _____ / _____

- 8.0 Invert and incubate the plates (30° - 35°C for *B. atrophaeus*; 55° - 60°C for *G. stearothermophilus*).

Incubation Start Time/Initial & Date: _____ / _____ Incubator #: _____

- 9.0 Examine all plates at 24 (±1) hours. Record on the back the number of colony forming units (CFU's) per plate. Record the average on the following page.
- 10.0 Calculate the average number of CFU's per carrier from the above data by using the formulas on the following page:

Performed By: _____ Date: _____

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Total @ 48 hrs / number of plates counted x DF = CFU/spore carrier
DF= Dilution factor (absolute value of the reciprocal of the dilution)
AV= Average number of colonies per spore carrier

Incubation End Time/Initial & Date: _____ / _____

CFU COUNTS AT 24 HOURS

dilutions _____

24hrs Plates 1. _____ 2. _____ 3. _____ Total @ 24hours: _____

Total @ 24 hrs _____ / 3 x _____ (DF) = _____ (AV)CFU/Spore carrier

CFU COUNTS AT 24 HOURS

dilutions _____

24hrs Plates 1. _____ 2. _____ 3. _____ Total @ 48hours: _____

Total @ 24 hrs _____ / 3 x _____ (DF) = _____ (AV)CFU/Spore carrier

of Dilutions = Dilution Factor

- 1 = 10
- 2 = 100
- 3 = 1000
- 4 = 10000
- 5 = 100000
- 6 = 1000000

Sum of the AV of both dilution / 2 =CFU/ Spore carrier

_____ / 2 =

_____ x 10^{_____} CFU/Spore Carrier

Read By: _____ Date: _____